

EFFECTS OF PHARMACOLOGICAL AGENTS ON THE ACTIVITY OF RAT KIDNEY ADENYLYL CYCLASE

Wadiah S. Backer¹ & Jalaluddin A. Khan²

ABSTRACT:

Objective: To study the effect of both selective and nonselective activator and inhibitory agent on adenylyl cyclase in rat kidney.

Design: Different concentrations of some pharmacological agents such as forskolin, nebularine, Ap₄A, Ap₃A and caffeine were prepared. The effects of the agents on the activity of rat kidney adenylyl were determined.

Methods: The crude extract obtained from rat kidney tissue was prepared and the specific activity of adenylyl cyclase in the crude extract was determined by using [2-H³] ATP as substrate which produced cAMP. Different concentrations of important pharmacological agents as activators and inhibitors of enzyme were examined. The compounds selected were forskolin as a known activator of adenylyl cyclase and for comparison; nebularine, Ap₄A, Ap₃A and caffeine were used.

Results: The results obtained showed that the highest activity of adenylyl cyclase was at 100μM forskolin. The activity of enzyme was inhibited by nebularine as the concentration of agent was increased until at 50μM where the inhibition began to level off. The effect of caffeine at 10-300μM on the activity of adenylyl cyclase showed no significant effect on the enzyme activity in kidney tissue. Ap₃A showed no effect on the activity of adenylyl cyclase over the concentration range of 10-300μM, whereas, Ap₄A produced an inhibition effect on the enzyme activity at concentration 100μM.

Conclusion: The important role of cyclic nucleotides in cell-signaling and metabolism control, evoke the activities and inhibitors of cyclase to play a potential physiological effects. As forskolin is well known activator of adenylyl cyclase, other pharmacological agents such as nebularine and Ap₄A are found to be a potential inhibitors of cyclase, which highlight these compounds as treatment of mania, schizophrenia, seizure and parkinson's diseases in which an increase of cAMP concentration has been demonstrated.

KEY WORDS: Nucleotides, Adenylyl cyclase, Forskolin, Nebularine, Ap₃A, Ap₄A, Caffeine.

Pak J Med Sci January-March 2004 Vol. 20 No. 1 41-45

1. Dr. Wadiah S. Backer
Dept. of Biochemistry, Girls Section
Faculty of Science, King Abdul Aziz University,
Jeddah, Saudi Arabia
2. Dr. Jalaluddin A. Khan
Dept. of Biochemistry
Faculty of Science, King Abdul Aziz University,
Jeddah, Saudi Arabia

Correspondence:

Dr. Wadiah Backer
P.O. Box: 80174,
Jeddah 21589, Saudi Arabia.
E-Mail: wadiahb@hotmail.com

- * Received for publication: April 30, 2003
Revision received: September 15, 2003
Revision accepted: September 17, 2003

INTRODUCTION

Because of the central role of cyclic nucleotides in cell-signaling and metabolic control, activators and inhibitors of the cyclases are potentially important as pharmacological agents. Some such compounds are already known and include forskolin. In this study, the activation produced by forskolin has been examined, and some other possible activators or inhibitors were examined for effect on kidney adenylyl cyclase. The effects of the compounds selected (nebularine, caffeine, Ap₃A, and Ap₄A) were compared to those of forskolin (a known activator of adenylyl cyclase). With the exception of forskolin, the compounds tested

were all purine derivatives, like cyclic AMP itself 'and ATP, the substrate of the enzyme'. Nebularine is a purine nucleoside antibiotic & has the structure purine 9- β -D-ribofuranoside. It is unique amongst the known naturally occurring purine ribosides in that it has no 6-substituent. The fungus *Lepista (Clitocybe) nebularis* produces the compound as a normal metabolite. It has high biological potency as a cytotoxin, inhibits the growth of seedlings of various species and causes mitotic aberrations in root tips. Also, it is an effective inhibitor of the multiplication of influenza B virus¹. In animal cells, nebularine causes mitotic aberrations, including chromosome breaks, and the compound is highly toxic to mice and guinea-pigs². The toxicity of the compound to animal cells in tissue culture is blocked by adenosine, AMP and ATP at concentrations 100-fold that of nebularine. Nebularine has also been reported to inhibit NAD⁺ - dependent glucose dehydrogenase and adenosine deaminase³.

The main source of caffeine is tea and coffee. It is a methylated xanthine and known to inhibit phosphodiesterases⁴.

Ap₃A and Ap₄A have been described as alarmones, analogous to hormones in that they regulate cell metabolism, and to alarms in that they signal when all is not well and help the cell adjust to its new conditions. They are diadenosine 5',5'''-P¹P⁴-tetrphosphate (Ap₄A) or its triphosphate (Ap₃A). Ames has proposed that Ap₃A represents, at least in bacteria, an alarmone that helps a cell respond to oxidative stress, imposed by agents such as hydrogen peroxide⁵. There is evidence that in animal cells Ap₄A serves as a positive regulator for DNA replication⁶. Both Ap₃A and Ap₄A are stored in large amounts in human platelets. After activation of the platelets, both of these dinucleotides are released into the extra cellular milieu where they play a role in the modulation of platelet aggregation and also in the regulation of the vasotone⁷. Similarly to Ap₃A, Ap₄A is degraded in rat plasma to ATP and AMP, and not to two ADP molecules, and these nucleotides are further degraded through adenosine⁸.

MATERIAL AND METHODS

Chemical and animal sources

All biochemicals including cyclic nucleotides and column chromatography materials were obtained from Sigma Chemical Co. (Poole, Dorset, U.K.). [2³H]adenosine-5'-triphosphate (ammonium salt) was obtained from Amersham International plc (Amersham, Buckinghamshire, U.K.). The high efficiency 'Ready Safe' scintillation cocktail, containing phenylxylylethane surfactant, used for aqueous samples was obtained from Amersham also.

The source of mammalian tissue used in this study was male Wister rats. The average weights were 250-350g. They were randomly selected from a rat colony bred from animals obtained in 1976 from Olac Ltd, U.K. and maintained in the experimental animal unit of King Fahad Medical Research Center, Jeddah. They were housed in an air-conditioned room at 24°C and maintained on a 12hr dark/light cycle. Animals were fed on standard food produced by Grain, Silos and Flour Mills Organization, Western Province Saudi Arabia.

Extraction of adenylyl cyclase activity

Weighed tissues samples were placed on ice and cut into small pieces, then homogenized in ice-cold 45mM Tris-HCl (pH 7.4) containing 6mM 2-mercapto-ethanol, 8mM theophylline and 0.25M sucrose for a total of 3min, in three 1 min bursts, at maximum speed (1500 rpm). For each g of tissue, 9ml of buffer were used. The homogenate was centrifuged at 3000g for 10min at 2°C. The precipitate was discarded, and the supernatant was recentrifuged at 150,000g for 40min at 2°C. The final precipitate was resuspended in 2-3ml of 45mM Tris-HCl (pH 7.4) containing 6mM 2-mercaptoethanol and 8 mM theophylline (in experiment of caffeine 8mM theophylline was not added to the Tris-HCl buffer to avoid having two methylated xanthines present in the same experiment). It was then divided into two parts, one was assayed for adenylyl cyclase activity and the other was used for determination of protein⁹.

Adenylyl cyclase assay

The procedure of Alvarez and Daniels¹⁰ was used with minor modifications. In the assay incubations, 20 μ l of the Tris-HCl buffer (pH 7.4) was replaced by 20 μ l of the possible activators or inhibitors solution of different concentrations.

RESULTS AND DISCUSSION

Forskolin is well known as an activator of adenylyl cyclase activity and in the present study its effect was maximal at 100 μ M (Fig. 1). This is similar to the effect previously reported for kidney tissue^{11,12}. The results, shown in Fig. 2, indicated a substantial inhibition by nebularine of adenylyl cyclase activity.

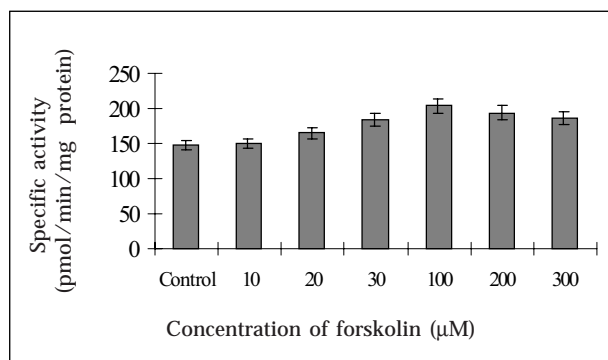


Figure-1: The stimulatory effect of forskolin on the adenylyl cyclase activity (pmol/min/mg protein) of rat kidney. 0.5mM [2- H^3] ATP was used as substrate. Incubation contained 4mM $MgSO_4$, 45mM Tris-HCl buffer at pH 7.4. Data points are each the mean of 12 replicate determination \pm SD.

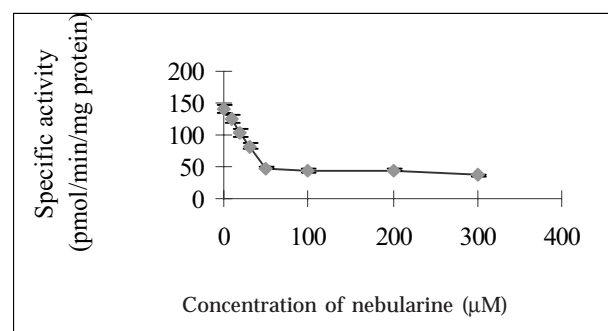


Figure-2: The inhibitory effect of nebularine on the adenylyl cyclase activity (pmol/min/mg protein) of rat kidney. 0.5mM [2- H^3] ATP was used as substrate. Incubation contained 4mM $MgSO_4$, 45mM Tris-HCl buffer at pH 7.4. Data points are each the mean of 12 replicate determination \pm SD.

Initially, as the nebularine concentration was increased, there was a sharp inhibition of activity until at 50 μ M nebularine, the inhibition was 66%. Between 50 and 300 μ M nebularine, the inhibition began to level off. A kinetic study using substrate (ATP) concentrations of 0.1 to 0.5mM, and inhibitor (nebularine) concentrations of 0, 10, 50 and 300 μ M showed that the inhibition was non-competitive (Fig. 3). This effect of nebularine on adenylyl cyclase activity is therefore similar to its effect on xanthine oxidase, a key enzyme of purine catabolism¹³. The inhibitory effect of nebularine on adenylyl cyclase activity could be due to an effect on the GTP-binding regulatory protein. For example, nebularine may prevent dissocia

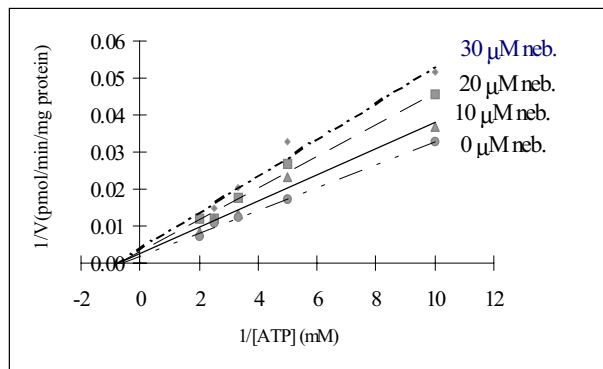


Figure-3: Lineweaver-Burk plot of adenylyl cyclase kinetics in the presence of nebularine. The enzyme was in 45mM Tris-HCl buffer (pH 7.4) containing 10mM theophylline. Data points are each of the mean of 12 replicate determination.

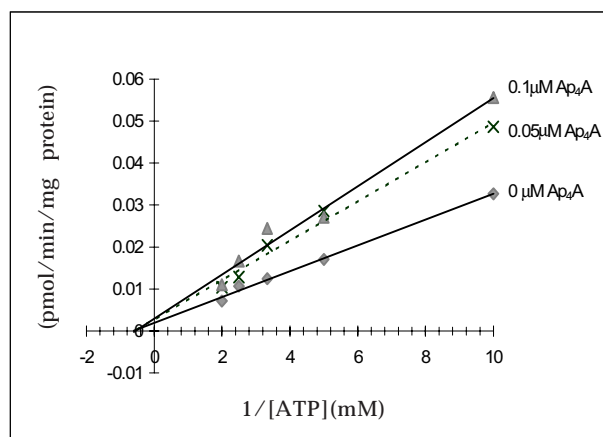


Figure-4: Lineweaver-Burk plot of adenylyl cyclase kinetics in the presence of Ap_4A . The enzyme was in 45mM Tris-HCl buffer (pH 7.4) containing 10mM theophylline. Data points are each of the mean of 12 replicate determination.

tion of the alpha subunit (α_s) of the stimulatory GTP-binding regulatory protein, G_s . Or nebularine may *in vivo* inhibit the hormone stimulating adenylyl cyclase by blocking the receptor.

The significant inhibition of adenylyl cyclase by nebularine highlights the potential importance of this compound both clinically and in the laboratory. It is possible for example, that nebularine could be of use in treating mania, schizophrenia, seizure, and Parkinson's disease, and other diseases in which an increase of cAMP concentration has been demonstrated. This compound could be considered as a selective inhibitor, since it has a high potency in inhibiting the activity of adenylyl cyclase.

The effect of caffeine was examined at 10-300 μ M on the activity of adenylyl cyclase (Table-I). The results showed no significant effect on the enzyme activity in kidney tissue. This result is in contrast to the reports of Sheppard¹⁴ and Jakobs *et al.*¹⁵ that methylxanthines in high concentrations inhibit adenylyl cyclase in some tissues. Furthermore, caffeine is known to inhibit the cAMP-induced activation of adenylyl cyclase by blocking signal transduction and not by acting directly on the enzyme¹⁶. Oyama and Kubota¹⁷ found that

TABLE-I: The effect of caffeine on adenylyl cyclase activity (pmol/min/mg protein) from rat kidney. The incubation mixture contained 0.5mM [2- H^3] ATP and 4mM $MgSO_4$. Kidney tissue was extracted with 45 mM Tris-HCl buffer (pH 7.4) without containing 10mM theophylline. Data are the mean of 5rats; each determination was in triplicate \pm SD

Caffeine (μ M) final concentration	Specific activity (pmol/min/mg protein)	Change relative to control (%)
0 (Control)	114 \pm 21	0
10	109 \pm 18	-5
20	113 \pm 19	-1
30	113 \pm 15	-1
100	112 \pm 16	-2
200	110 \pm 13	-4
300	114 \pm 17	-0

caffeine inhibited the cation-induced activation of adenylyl cyclase in the saponin-treated cells, of the cellular slime mold *Dictyostelium discoideum*. Stimulation of striatal adenylyl cyclase by forskolin is not however significantly affected after chronic caffeine ingestion in mice¹⁸.

This result indicates that the daily intake of caffeine in drinks such as tea, coffee, and cola

TABLE-II: The effect of Ap_3A on adenylyl cyclase activity (pmol/min/mg protein) of rat kidney. The incubation mixture contained 0.5mM [2- H^3] ATP and 4mM $MgSO_4$. Kidney tissue was extracted with 45mM Tris-HCl buffer (pH 7.4) containing 10mM theophylline. Data are the mean of 5rats; each determination was in triplicate \pm SD

Ap_3A (μ M) final concentration	Specific activity (pmol/min/mg protein)	Change relative to control (%)
0 (Control)	112 \pm 15	0
10	112 \pm 11	0
20	113 \pm 14	-1
30	103 \pm 12	-8
100	104 \pm 13	-7
200	107 \pm 6	-5
300	107 \pm 8	-5

TABLE-III: The inhibitory effect of Ap_4A on adenylyl cyclase activity (pmol/min/mg protein) of rat kidney. The incubation mixture contained 0.5mM [2- H^3] ATP and 4mM $MgSO_4$. Kidney buffer (pH 7.4) containing 10mM theophylline. Data are the mean of 5 rats; each determination was in triplicate \pm SD

Ap_4A (μ M) final concentration	Specific activity (pmol/min/mg protein)	Change relative to control (%)
0 (Control)	125 \pm 15	0
10	117 \pm 7	-6
20	115 \pm 13	-8
30	104 \pm 6	-17
50	98 \pm 4	-22
100	81 \pm 5	-35
200	146 \pm 17	+17
300	133 \pm 18	+6
1000	123 \pm 12	-2

does not affect the biochemical machinery of kidney metabolism but this does not necessarily mean that high intakes of caffeine is harmless.

Ap₃A and Ap₄A were added, separately, to adenylyl cyclase incubation mixtures. Ap₃A had no effect on the activity of adenylyl cyclase over the concentration range of 10-300 μM (Table-II), whereas, Ap₄A at a concentration of 100 μM, inhibited the activity of adenylyl cyclase by 36% of the control value (Table-III). The Lineweaver-Burk plot, of the substrate ATP concentration 0.1-0.5 mM and Ap₄A at concentration 0.05 and 0.1 mM shown in Fig. 4 indicates that Ap₄A is a non-competitive inhibitor of adenylyl cyclase. This results suggested that Ap₄A could be used, similarly to nebularine, for treatment of diseases in which cAMP accumulates and as an 'alarmone' through inhibiting a cyclic nucleotide system.

REFERENCES

1. Tamm I, Folkers K & Shunk CH. A certain benzimidazoles, benzenes, and ribofuranosylpurines as inhibitors of influenza B virus multiplication, *J Bacteriol.* 1956; 72: 59-62.
2. Truant A P & D'amato H E. Pharmacologic and toxicologic actions of nebularine, *Fed Proc.* 1955; 14: 391-5.
3. Nair V & Weichert RJ. Substrate specificity of adenosine deaminase-function of the 5'-hydroxyl group of adenosine, *Bioorg. Chem.* 1980; 9: 423-7.
4. Drummond GI & Duncan L. Adenyl cyclase in cardiac tissue, *J. Biol. Chem.* 1970; 245: 976-83.
5. Bochner BR., Lee PC, Wilson SW, Cutler CW & Ames BN. AppppA and related adenylylated nucleotide are synthesized as a consequence of oxidation stress, *Cell* 1984;37: 225-32.
6. Zamecnik PC. Diadenosine 5',5'''- P1, P4-tetraphosphate (Ap₄A): its role in cellular metabolism, *Anal. Biochem.* 1983; 134: 1-10.
7. Luthje J. & Ogilvie A. Catabolism of Ap₄A and Ap₃A in whole blood, *Eur J Biochem.* 1988; 173: 241-5.
8. Kimura T, Hatano N, Wada M, Iwata K, Kurosaki Y, Nakayama T, Yamaura T & Nakajima H. Disposition of diadenosine 5',5'''- P1,P4-tetraphosphate (Ap₄A) in rats, *Biol Pharm Bull.* 1995; 18(11): 1556-9.
9. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding, *Anal Chem.* 1976; 72: 248-54.
10. Alvarez R & Daniels DV. A separation method for the assay of adenylyl cyclase, intracellular cyclic AMP, and cyclic-AMP phosphodiesterase using tritium-labeled substrates, *Anal Biochem.* 1992; 203: 76-82.
11. Takats A, Binh VH & Bertok L. Potential role of SH group in the radiosensitivity of adenylate cyclase, *Acta Physiol Hung.* 1990; 76(4): 265-72.
12. Marano I, Adler K, Weismann K, Knorr A, Erdmann E & Bohm M. Correlation of myosin heavy chain expression in the rat with cAMP in different models of hypertension-induced cardiac hypertrophy, *J Mol Cell Cardiol.* 1993; 25(4): 387-94.
13. Brown E & Konuk M. Plant cytotoxicity of nebularine (purine riboside), *Phytochemistry.* 1994; 37(6): 1589-92.
14. Sheppard H. Inhibition of norepinephrine stimulated adenyl cyclase by theophylline, *Nature (London)* 1970; 228: 567-71.
15. Jakobs KH, Schultz K & Schultz G. Inhibition of adenyl cyclase preparations from rat kidney by calcium ions and various diuretic, *Nauryn. Schmiedeberg's Arch Pharmacol.* 1972; 273: 248-55.
16. Brenner M & Thoms SD. Caffeine blocks activation of cyclic AMP synthesis in *Dictyostelium discoideum*, *Dev Biol.* 1984; 101:136-46.
17. Oyama M & Kubota K. Activation of adenylate cyclase by divalent cations and polyamines in saponin-treated *Dictyostelium discoideum* cells, *J Biochem. Tokyo*, 1995; 118(1): 117-21.
18. Shi D, Nikodijevic O, Jacobson KA & Daly JW. Effects of chronic caffeine on adenosine, dopamine and acetylcholine systems in mice, *Arch Int Pharmacodyn Ther.* 1994; 328(3): 261-87.